

# *Cryptococcus neoformans* var. *grubii*: The *In Vitro* Antifungal Susceptibility Pattern in Addition to the Quantification of Phospholipase and Proteinase Enzymatic Activities

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## Article history:

Received: January 18, 2022

Accepted: March 25, 2023

Published: March 29, 2023

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## Abstract

**Background:** Cryptococcal meningoencephalitis is a life-threatening fungal infection in human immunodeficiency virus (HIV)-infected patients. *Cryptococcus neoformans* var. *grubii* and *neoformans* are the causative agents that usually respond well to fluconazole and amphotericin B. However, resistance/non-responding cryptococcal meningitis cases to fluconazole and amphotericin B have been reported globally.

**Methods:** The causative *Cryptococcus* was identified by phenotypic and singleplex polymerase chain reaction (PCR) targeting the putative sugar transporter (*STR1*) gene. In addition, the phospholipase and proteinase enzymatic activities of the isolates were determined by the plate method using egg yolk agar and bovine serum albumin agar plates, respectively. Finally, the in-vitro minimal inhibitory concentration (MIC) of fluconazole, voriconazole, and amphotericin B against isolated *C. neoformans* strains was determined by the broth microdilution method.

**Results:** A total of 50 *C. neoformans* strains were isolated from the cerebrospinal fluid of HIV-infected patients, which were further identified as variety *grubii* by simplex polymerase chain reaction (PCR). All the isolated strains producing phospholipase and proteinase enzymes were determined by the calculation of Pz, a ratio of colony diameter and diameter of colony plus the precipitation zone. A comparative high proteinase enzyme activity was observed, and these strains produced medium to high phospholipase (mean Pz  $0.3720 \pm 0.082$ , range 0.23-0.56) and proteinase activity (Mean Pz  $0.3069 \pm 0.086$ , range 0.012-0.54). A varied antifungal MIC was detected, and voriconazole had the lowest MIC<sub>50</sub> and MIC<sub>90</sub> (0.03 & 0.06 µg/mL) in comparison to fluconazole and amphotericin B.

**Conclusion:** *Cryptococcus neoformans* var. *grubii* is the commonest cause of cryptococcal meningoencephalitis in HIV-infected patients. The isolates had varied extracellular hydrolytic enzyme activities. The emergence of *C. neoformans* strains with higher fluconazole MIC ( $\geq 4$  mcg/mL) could have resulted in treatment failure.

**Keywords:** Amphotericin B, Fluconazole, Var. *grubii*, HIV, *str1* gene, MIC



**Please cite this article as follows:** Gupta MK, Tilak R, Pal N, Singh AK, Chakravarty J, Kumar B. *Cryptococcus neoformans* var. *grubii*: The *In Vitro* antifungal susceptibility pattern in addition to the quantification of phospholipase and proteinase enzymatic activities. Avicenna J Clin Microbiol Infect. 2023; 10(1):32-37. doi:10.34172/ajcmi.2023.3367

## Introduction

*Cryptococcus neoformans* is a common cause of meningoencephalitis in human immunodeficiency virus (HIV)-infected patients (1). Two species of *Cryptococcus* named *neoformans* and *gattii* cause human infections. *C. neoformans* usually causes meningoencephalitis in immunocompromised patients, whereas *C. gattii* is manifested as cryptococcoma in immunocompetent individuals. Based on the serotypes whether A or D,

*C. neoformans* is further distinguished as two varieties called *grubii* and *neoformans*. Among these two, *grubii* is reported to be the most common cause of cryptococcal infections in immunocompromised individuals (2).

*Cryptococcus neoformans*, an environmental saprotroph, enters the human body through inhalation and results in self-limiting localized respiratory infections in the immunocompetent individual as the alveolar macrophages not only phagocytose the inhaled yeast



cells promptly but also recruit other inflammatory cells, viz. neutrophils, macrophages, and natural killer cells at the site of infection. However, in immunocompromised patients, HIV-infected patients having CD4 + T cell < 100 cells/ $\mu$ L, patients on chemotherapy, radiotherapy, corticosteroids, and the like, *Cryptococcus* escapes the immune system and disseminates hematogenously where it involves the distant body organs, brain parenchyma, genitourinary tract, and skin (1, 3). Different factors such as polysaccharide capsule, extracellular enzymes (e.g., urease, proteinase, phenoloxidase, and phospholipase), and the ability to grow at 37°C accentuate its adhesion, invasion, dissemination, and survival from the innate immune system (4). The identification of such factors helps in understanding its pathogenesis.

Different groups of antifungal agents, including azoles, polyene, and antimetabolites, are used to treat cryptococcal meningitis. Fluconazole and amphotericin B are widely employed in developing countries. However, *C. neoformans* strains, resistant/non-responding to amphotericin B, fluconazole, and 5-flucytosine have been reported around the globe, resulting in treatment failure/relapse (5-7). Keeping these facts in mind, a prospective study was performed to determine the variety of *Cryptococcus* causing human infections and their antifungal susceptibility for fluconazole, amphotericin B, and voriconazole. In addition, the study quantified the extracellular phospholipase and proteinase hydrolytic enzyme activities.

### Materials and Methods

The prospective study was approved by the Ethics Committee of the Institute of medical Sciences, BHU Varanasi (Dean/2019/EC/1021). *Cryptococcus* strains were isolated from the cerebrospinal fluid of HIV-infected patients, anti-retroviral therapy (ART) Clinic, Sir Sundarlal Hospital, Institute of Medical Sciences, Banaras Hindu University, Varanasi, India.

We isolated and identified the yeast causing meningoencephalitis in HIV-infected patients, presented to the ART Clinic, Sir Sundarlal Hospital, Institute of Medical Sciences, Banaras Hindu University, Varanasi, India, as per the standard mycology procedures, including the capsule demonstration by India ink wet mount, culture characteristics, urea hydrolysis test, and the growth at 37°C. The isolated *Cryptococcus* was speciated by the singleplex polymerase chain reaction (PCR) using the primers targeted to the putative sugar transporter (*STR1*) gene. Briefly, the cryptococcal strains were cultivated on the yeast extract peptone dextrose (YPD) broth, Himedia, India medium overnight. The cells were centrifuged and suspended in 400  $\mu$ L lysis buffer containing TE buffer and sodium dodecyl sulfate. The suspension was boiled twice with intermittent vortexing. Subsequently, lyticase was added to break the fungal cell wall. The CTAB extraction buffer was then added, and purification with phenol [chloroform: Isoamyl alcohol

(25:24:1)] was performed after incubation at 65°C. Finally, the extracted DNA was precipitated with chilled isopropanol. DNA was dissolved in the TE buffer (pH: 8.6), and the quantity of the extracted DNA was measured by a nanodrop spectrophotometer. Next, the putative sugar transporter (*str1*) gene was amplified by using *str1F* (5'GAGATTCGGCAGGAAGAAGC3') and *str1R* (5'CGTAAGGGATGACGAAAAGGTA3') primers in a reaction volume of 25  $\mu$ L, having 2.5  $\mu$ L of 10X buffer, 2.5  $\mu$ L of 10 nM dNTP mixture, 1 unit Taq Polymerase, and 1  $\mu$ L each primer with 5  $\mu$ L of extracted DNA.<sup>8</sup> Then, thermal conditions were followed for the PCR for 30 cycles of amplification, including initial denaturation at 94°C for 5 minutes, annealing at 55°C for 30 seconds, and elongation at 72°C for 30 seconds with a final extension step at 72°C for 6 minutes. The amplicon was subsequently electrophoresed on 1.2% agarose gel, and the results were compared with previous reports (8).

All the isolated strains were assessed for polysaccharide capsule, urease production, growth at 37°C, and the presence of the phenoloxidase enzyme by standard mycological procedures. Phospholipase and proteinase activities were quantified by the plate method (9,10). The Sabouraud dextrose agar medium supplemented with 1 mol/L sodium chloride, 0.005 mol/L calcium chloride, and 8% sterile egg yolk suspension was used to determine the phospholipase activity, whereas bovine serum albumin (BSA) agar containing yeast carbon base (1.17%), 0.2% BSA, and 2% agar was employed to quantitate the proteinase enzyme (9,10). The 10  $\mu$ L thick suspension of the isolated strains was placed in the centre of the above-mentioned media. The plates were incubated at 37°C for five successive days in a biochemical oxygen demand incubator. The tests for enzyme activities were executed in triplicates where an average value of the colony diameter (*a*) and the diameter of the colony plus precipitation zone (*b*) underwent measurement. The enzyme activities of these isolated strains were expressed as Pz value (*a/b*) as described by Price et al (11). According to this definition, a low Pz value implies high enzyme production, while a high Pz value indicates low enzyme production. All the isolated strains were grouped into different classes such as high (Pz < 0.4), medium (Pz 0.41-0.60), low (Pz 0.61-0.80), very low (Pz 0.81-0.99), and none (Pz 1) according to previous research (12).

The antifungal susceptibility of fluconazole, amphotericin B, and voriconazole against isolated *Cryptococcus* was executed by the broth microdilution method using the Clinical and Laboratory Standards Institute (CLSI) M27A<sub>3</sub> guidelines. Briefly, 0.5 McFarland suspension of the strains having a concentration of 0.5-1.5  $\times 10^6$  cells/mL was prepared, which was further diluted 20 times in RPMI1640 and 50 times in normal saline to have a final concentration of 0.5-2.5  $\times 10^3$  yeast cells/mL. Fluconazole, voriconazole, and amphotericin B drug powder were procured from Sigma Pharmaceuticals, India. Fluconazole was dissolved in distilled water to make

a master stock of 6.4 mg/mL, whereas voriconazole and amphotericin B were dissolved in DMSO to make a master stock of 1.6 mg/mL. These master stock solutions were further diluted (1:50) in RPMI 1640 to make a working solution of 128 µg/mL for fluconazole and 32 µg/mL for amphotericin B and voriconazole. These dilutions were prepared by two-fold drug dilution using RPMI1640 broth (13).

*Candida parapsilosis* ATCC 22019 and *Candida krusei* ATCC 6258 strains were used to standardize the broth microdilution assay. RPMI 1640 broth, without bicarbonate, buffered to a pH rate of 7.0 with morpholinepropanesulfonic acid was employed to determine the minimum inhibitory concentration of the above-mentioned drugs. Microdilution assay was performed in 96-well, flat-bottom microtitre plates in which 100 µL volume of the diluted drug and 100 µL of yeast suspension were placed, and these plates were incubated at 35°C for 72 hours and examined for growth. For amphotericin B, the minimal inhibitory concentration (MIC) was determined where a 100% growth reduction was observed, whereas 80% growth inhibition was noted as the MIC for fluconazole, and voriconazole compared to the growth control (13).

### Statistical Analysis

The distributions of datasets obtained in the study were checked for normality using the Kolmogorov-Smirnoff test. The means were separated using Tukey's test when data were normally distributed, and variances were homogeneous (Bartlett's test for equal variances). The data were presented as the mean ± standard deviation (SD), along with their confidence interval values. Student's t-test was conducted to analyze and compare the phospholipase and proteinase enzyme activities of *C. neoformans*. However, the chi-square test was performed for the percent analysis of the performance of both enzymes. The obtained *P* values less than 0.05 were considered statistically significant. All statistical analyses were conducted using SPSS software, version 23.0.

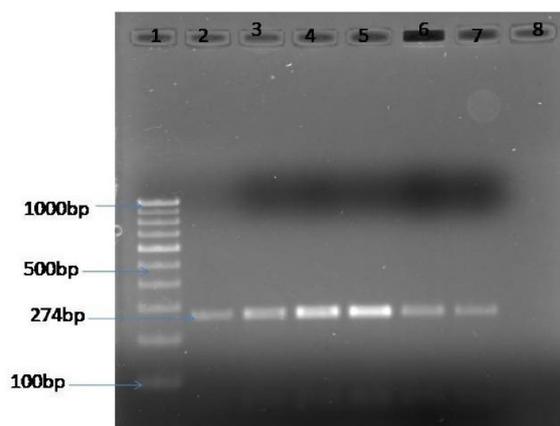
### Results

A total of 50 *Cryptococcus* strains were isolated from the cerebrospinal fluid of HIV-infected patients in 2 years. These strains were identified by standard mycology procedures, including culture characteristics, urease enzyme production, and growth at 37°C. Further characterization of these *Cryptococcus* strains was performed by the singleplex PCR, targeting the *STR1* gene, which revealed *C. neoformans* var. *grubii*, a sole causative agent in all 50 cases (Figure 1). All these 50 strains determined virulence factors such as the polysaccharide capsule, urease enzyme, phenoloxidase enzyme, phospholipase, and proteinase enzyme.

The quantification of the phospholipase and proteinase enzyme activities of the isolated *C. neoformans* strains was done by the plate method. A significantly high proteinase

enzyme activity was observed compared to phospholipase activity (t-value=3.874 and *P*=0.0002). All 50 *C. neoformans* strains had medium to strong phospholipase activity and proteinase enzyme activities with Pz value [mean (±SD): 0.3720 (0.082), range 0.23-0.56] and [Mean (±SD) 0.3069 (±0.086), range 0.012-0.54], respectively (Table 1). Moreover, when enzyme activity was categorized into strong and medium types by taking the cutoff enzyme activity at the Pz value of ≤0.4 and 0.41-0.60 as strong and medium and analyzed, a significantly stronger proteinase enzymatic activity was found than that of phospholipase (Table 2).

The MIC of fluconazole, voriconazole, and amphotericin B against the isolated *C. neoformans* strains was determined by the broth microdilution method. The MIC of the tested antifungals against *C. parapsilosis* ATCC 22019 and *C. krusei* ATCC 6258 was within the susceptibility range. A varied antifungal MIC was observed against the tested strains (Table 3). A higher fluconazole MIC than that of amphotericin B and voriconazole was observed as well. The mean, MIC<sub>50</sub>, and MIC<sub>90</sub> of fluconazole were 1.0175 ± 0.799, 1, and 2 µg/mL; whereas 0.0455 ± 0.029, 0.03, and 0.125 µg/mL were the mean, MIC<sub>50</sub>, and MIC<sub>90</sub> of voriconazole against all 50 isolates. The 0.1643 ± 0.131,



Lane1: 1Kb ladder; Lane 2 to 7: sample 1,2,3,4,5,6; Lane 8: Negative control

Figure 1. Gel Electrophoresis Image Showing the Band Size of The Polymerase Chain Reaction Products of *Cryptococcus neoformans*

Table 1. Comparison of Mean Phospholipase and Proteinase Enzyme Activities Produced by Isolated *Cryptococcus neoformans* var. *grubii* Strains

Enzyme Activity	Mean Pz Value	Standard Deviation	95% CI	t-value	P Value
Phospholipase	0.3720	0.82	-0.0984 to	-3.874	0.0002
Proteinase	0.3069	0.086	-0.0318		

Note. CI: Confidence interval.

Table 2. Comparison of Phospholipase and Proteinase Enzyme Activities of *Cryptococcus neoformans* var. *grubii* Strains

Enzyme Activity	Strong No. (%)	Medium No. (%)	Total	Chi-square Value	P Value
Phospholipase	30 (60%)	20 (40%)	50	10.1871	0.001414
Proteinase	44 (88%)	6 (12%)	50		

0.125, and 0.25 µg/mL were also observed as the mean, MIC<sub>50</sub>, and MIC<sub>90</sub> of amphotericin B against all isolates (Figure 2, Table 4).

**Discussion**

*Cryptococcus neoformans* is the opportunistic yeast, which is manifested as meningoencephalitis among HIV-infected patients having CD4+T cells < 100/µL. *C. neoformans* is divided into three serotypes, A, D, and AD hybrid. Serotype A is known as *grubii*, whereas serotype D is known as *neoformans*. These two varieties of *C. neoformans* primarily result in these central nervous system (CNS) manifestations worldwide. The *grubii* variety is the most common cause of about 95% of cases of cryptococcal infection worldwide, whereas the *neoformans* variety causes ~5% of the cryptococcal infection in western countries (3). In the present study, *grubii* (n = 50) is the sole causative agent of all cryptococcal meningitis cases among HIV-infected patients. A similar study in North India has reported *C. neoformans* var *grubii* as a causative agent in all 46 cases of cryptococcal meningitis (14). The causative variety is identified by phenotypic, serological, and molecular methods. Phenotypic and serological methods are time-taking and laborious; thus, this differentiation can be executed by the singleplex PCR targeting the *str1* gene by which we differentiate the two varieties of the *C. neoformans* and identify the *C. gattii* species (8).

Being environmental saprotrophs, the basidiospores of *Cryptococcus* aerosolize and deposit in the host alveoli, where active cell-mediated immunity impedes transmission. However, in HIV-infected patients and

immunocompromised individuals, the virulence factors of *Cryptococcus*, the polysaccharide capsule, phenoloxidases, urease, phospholipase, and proteinase act together and lead to a disease state by mechanisms such as adhesion, invasion, and dissemination (2). The polysaccharide capsule and urease enzyme production are the major virulence factors of *Cryptococcus*. The capsule prevents phagocytosis, whereas the urease enzyme helps in the intracellular survival of *C. neoformans* by increasing pH (4,15). Nonetheless, the phenoloxidase enzyme makes it a neurotrophic yeast (2,4). Enzymes urease, phenoloxidase, proteinase, and phospholipase were produced by all 50 isolated strains, but in the present study, we only quantified phospholipase and proteinase enzymes.

The phospholipase enzyme demonstrates phospholipase B, lysophospholipase hydrolase, and lysophospholipase transacylase activities which support fungal cell attachment. This enzyme also degrades the phospholipid component of the cell membrane, accentuating its invasion (16). Cryptococcal proteinase enzyme degrades different cellular proteins, including collagen, elastin, fibrinogen, complement factors, and immunoglobulin, disrupting the host tissue barriers (17). Thus, all these enzymes, along with the capsule, accentuate the pathogenesis. The roles of phospholipase and proteinase enzymes in the invasion and dissemination of *Cryptococcus* have also been reported in other studies (16-18).

Cryptococcal meningoencephalitis cases are managed by combination therapy, including amphotericin B, fluconazole, and 5-flucytosine. Amphotericin B binds to the sterol component of a fungal cell wall, resulting in pore

**Table 3.** Number of *C. neoformans* var. *grubii* Strains Having Specific Antifungal MIC

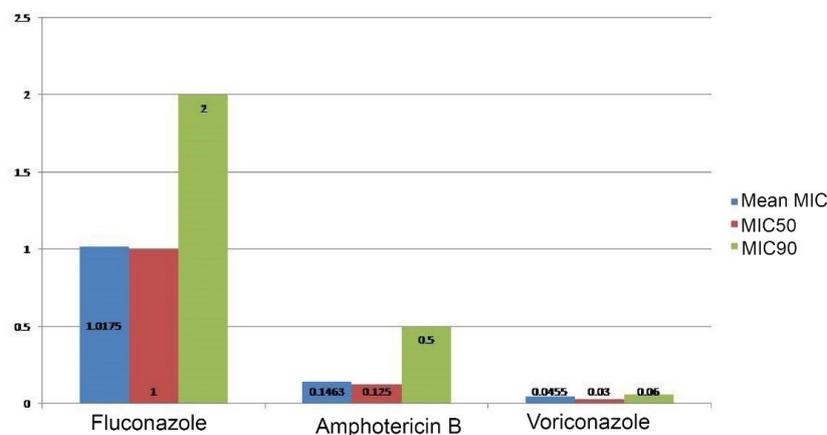
Drug	MIC (µg/mL)											
	0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64
Fluconazole	0	0	1	3	18	19	7	2	0	0	0	0
Voriconazole	35	10	5	0	0	0	0	0	0	0	0	0
Amphotericin B	4	12	19	10	5	0	0	0	0	0	0	0

Note. MIC: Minimal inhibitory concentration.

**Table 4.** Mean, MIC<sub>50</sub>, and MIC<sub>90</sub> of Tested Antifungal Agents Against *Cryptococcus neoformans* var. *grubii* Strains

Antifungal Agent	MIC Range (µg/mL)	MIC <sub>50</sub> (µg/mL)	MIC <sub>90</sub> (µg/mL)	Mean MIC ± SD
Fluconazole	0.125-4	1	2	1.017 ± 0.79
Voriconazole	0.03-0.125	0.03	0.06	0.0455 ± 0.029
Amphotericin B	0.03-0.5	0.125	0.25	0.1463 ± 0.114

Note. MIC: Minimal inhibitory concentration; SD: Standard deviation.



**Figure 2.** Mean, MIC<sub>50</sub>, and MIC<sub>90</sub> of Antifungal Agents Against Isolated *Cryptococcus neoformans* Strains. Note. MIC: Minimal inhibitory concentration

formation with subsequent lysis. In contrast, fluconazole inhibits the synthesis of ergosterol, an important component of the fungal cell membrane. In addition, 5-fluorocytosine inhibits the synthesis of fungal nucleic acid. *C. neoformans* strains usually respond well to the above-mentioned drugs. However, fluconazole resistance/non-susceptibility/increasing MIC has been reported from different parts of the globe (5,6,7). Inadequate doses of fluconazole with widespread use in agriculture as a preventive tool for black banana rot disease may attribute the emergence (19). ARTEMIS DISK Global Antifungal Surveillance Study has reported constantly increasing fluconazole resistance among *C. neoformans* from 7.3% to 11.7% (20). Non-responding/increasing fluconazole MIC against *C. neoformans* strains has been reported from Taiwan and Uganda. In the present study, 1 and 2 µg/mL fluconazole MIC<sub>50</sub> and MIC<sub>90</sub> were observed, respectively, against 50 isolates of *C. neoformans*, whereas it was 8 and 32 µg/mL in Uganda (21). Our results are in line with the findings of a study by Chaudhary et al, reporting 2 and 4 µg/mL fluconazole MIC<sub>50</sub> and MIC<sub>90</sub>, respectively, against the isolated *C. neoformans* var. *grubii* strains (22). Another study from AIIMS, New Delhi reported a higher fluconazole MIC<sub>50</sub> and MIC<sub>90</sub> values of 4 and 16 µg/mL, respectively (23). This variability may be attributed to the AIIMS, an apex centre of health care in India, where the most referred patients are treated.

Fluconazole is widely used in the management of cryptococcal meningoencephalitis. For effective management, the adequate drug concentration is to be achieved at the site of action (brain parenchyma). However, a lower fluconazole cerebellum AUC/plasma AUC of 46.9% has been reported earlier by Sudan et al (24,25). Thus, conventional fluconazole doses can easily tackle the strains with a low fluconazole MIC. In contrast, strains with a higher MIC have to be managed either by high drug doses or the drug combination. Accordingly, treatment failure is a common occurrence with a higher fluconazole MIC, as strains having a lower fluconazole MIC are eliminated at conventional dosing, leaving behind the subpopulation with a higher MIC, subsequently resulting in treatment failure/relapse (26,27). CLSI recommends an 8 µg/mL fluconazole epidemiological cutoff against *C. neoformans* (28). However, Sudan et al reported 2 µg/mL MIC as a fluconazole breakpoint against *C. neoformans* isolated from meningoencephalitis cases (24).

Voriconazole has a similar mechanism of action to fluconazole. Espinel-Ingroff et al reported epidemiological cutoff (ECOFF) for voriconazole (0.25 µg/mL) against *C. neoformans* and *C. gattii* (28). Our study results revealed a lower (0.03 µg/mL) voriconazole MIC<sub>50</sub>. Conversely, Thompson et al found a higher voriconazole antifungal activity against *C. neoformans* strains (29). Thus, a lower voriconazole MIC against isolated *C. neoformans* may provoke clinicians for its use in treatment as it can easily cross the blood-brain barrier.

Amphotericin B binds the sterol component of

the fungal cell wall, resulting in pore formation with subsequent leakage of cytoplasmic content and cell death. Therefore, amphotericin B, along with fluconazole, is used in the continuous phases of the treatment. Espinel-Ingroff et al reported (0.5 µg/mL) an epidemiological cutoff for amphotericin B against *C. neoformans* (28). All the isolated strains had a MIC within epidemiological cutoff values in our study. These results conform to the findings of the study by Chaudhary et al, demonstrating 0.235, 0.25, and 0.5 µg/mL amphotericin B mean, MIC<sub>50</sub>, and MIC<sub>90</sub> values, respectively, against 160 clinical isolates of *C. neoformans* var. *grubii* (22).

Although the MIC of the tested antifungal agents is within the epidemiological cut-off, a close watch on the treatment response is the need of time as in the last decade. Numerous cases of relapse/ treatment failure have been reported globally. Sudan et al found a higher 96.8% success rate in patients infected with *C. neoformans* with 2 µg/mL fluconazole MIC compared to a 52% success rate in patients infected with strains having 4 µg/mL MIC (24). However, in the present study, we only determined the in-vitro MIC of fluconazole, voriconazole, and amphotericin B against isolated *Cryptococcus* strains, without any correlation of clinical responses with the antifungal MIC. Thus, there is a need for a prospective study to determine the correlation of fluconazole and amphotericin B MICs with treatment outcomes in cryptococcal meningoencephalitis.

## Conclusion

*Cryptococcus neoformans* var. *grubii* is the sole causative agent of meningoencephalitis among enrolled patients. All the isolates have urease, proteinase, and phospholipase enzymes. Moreover, the emergence of *C. neoformans* strains having a higher fluconazole MIC might result in treatment failure/relapse in meningoencephalitis cases.

## Authors' Contribution

**Conceptualization:** Munesh K Gupta, Ragini Tilak, Jaya Chakravarty.

**Data curation:** Munesh K Gupta, Namrata Pal, Bhupendra Kumar.

**Formal analysis:** Munesh K Gupta, Namrata Pal, Ashish Kumar Singh, Bhupendra Kumar.

**Investigation:** Munesh K Gupta, Namrata Pal, Ashish Kumar Singh.

**Methodology:** Munesh K Gupta, Namrata Pal, Ragini Tilak, Jaya Chakravarty, Bhupendra Kumar, Ashish Kumar Singh.

**Resources:** Munesh K Gupta, Ragini Tilak, Jaya Chakravarty.

**Software:** Ashish Kumar Singh, Bhupendra Kumar, Munesh K Gupta.

**Supervision:** Ragini Tilak, Jaya Chakravarty, Munesh K Gupta.

**Validation:** Ragini Tilak, Munesh K Gupta.

**Visualization:** Ragini Tilak, Jaya Chakravarty.

**Writing—original draft:** Namrata Pal, Ashish Kumar Singh, Bhupendra Kumar, Munesh K Gupta.

**Writing—review & editing:** Munesh K Gupta, Ragini Tilak, Jaya Chakravarty, Bhupendra Kumar.

## Competing Interests

There is no conflict of interest.

## Funding

There is no source of funding.

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